

GLOBAL USE OF ANTIMICROBIALS IN FOOD ANIMALS, EMERGENCE OF ANTIMICROBIAL RESISTANCE AND WAY FORWARD: AN OVERVIEW

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Several decades of antibiotic abuse in humans, animals and other farming systems and release of antimicrobials into environment from different sources are contributing to development of resistance to multiple classes of antibiotics in bacteria at a rate much faster than the rate at which newer antibiotics are being developed thus creating health emergency situation. Use of antibiotics as growth promoters (AGPs), wherein antibiotics are used at sub therapeutic dose for longer duration, favours selection and spread of resistant bacteria within animals and to humans through food or other environmental pathways. Expressed concerns are that AGPs are used to compensate for poor hygiene, health management and housing. Considering gravity of threat to human and animal health, some countries have banned use of AGPs while many countries are still using various antimicrobials as AGP including some of those categorized as important for human health. Recently many antibiotics being used in animals for therapeutic purpose are also being implicated for development of resistance. Different alternatives to AGPs are available but they may not be as cheaper as and also may not be as effective as antibiotics for improving survivability or performance of food animals reared under poor hygienic conditions or under environmental stress. Data on prevalence of antimicrobial resistance (AMR) is limited in India. There is urgent need to undertake detail study of prevalence of AMR and economic analysis of rearing food animals on various alternatives to antibiotics. Further, development of reliable certification system and higher pricing of animal products produced through alternate system and increasing consumer awareness can be a way forward to tackle the menace.

Key words: Alternatives to antibiotics, Antibiotic growth promoters, Antimicrobial resistance

Probably antibiotics have been around for millions of years as an outcome of mechanism of struggle for survival of microorganisms in hostile environment although scientific community came to know about their existence only in 1928 with discovery of the first antibiotic named as penicillin by Alexander Fleming and the research was

formally published in the year 1929 (Fleming, 1929). Within four years of introduction of the first antibiotic penicillin for therapeutic use infections resistant to the drug began to appear. Emergence of antimicrobial resistance (AMR) is not unexpected considering that bacteria have been around for nearly four billion years and evolving in

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very hostile environments, an essential characteristic for survival would have been the capacity to deal with harmful chemicals like antibiotics released by other bacteria and fungi (Rodríguez-Rojas *et al.*, 2013; Blair *et al.*, 2015). In a study, DNA isolated from 30,000-year-old permafrost cores, resistance genes to β -lactams, tetracycline and vancomycin were found (D'Costa *et al.*, 2011). The 1940-1962 remained the golden era of antibiotic discovery. Many of the classes of antibiotics used today were discovered during this period. Presently only few newer antibiotics are under development. Unregulated use and misuse of antibiotics are giving rise to antibiotic resistance at faster rate than the rate at which newer antibiotics are being developed. Available reports have estimated a per annum human death toll of 23,000 in the USA, 25,000 in EU, and 58,000 in India from various drug resistant bacterial infections (Chaudhary, 2016). Thus emergence of antimicrobial resistance has become a global health challenge and major threat to human and animal health. AMR affects livestock productivity, impacting farmer livelihoods, food security and safety. In India, there is increasing evidence of resistance in human and animal pathogens to both older antimicrobials and newer critically important drugs like carbapenems (Kumaraswamy *et al.*, 2010). Despite this, India has a paucity of reliable data on the quantities of, and drivers for antimicrobial use in humans and livestock production.

Global use of antibiotics in food animals

The amount of antimicrobials used in food animals has been estimated to escalate globally from 131,209 tons in 2013 to 200,235 tons by 2030 - an increase of 53% (Van Boeckel *et al.*, 2017). The followings

are the estimated global average annual consumption of antimicrobials to produce one kilogram of meat: 45 mg/kg beef, 148 mg/kg chicken and 172 mg/kg pork (Van Boeckel *et al.*, 2015). China, USA, Brazil, India and Germany utilize 23, 13, 9, 3 and 3 % of global antimicrobial use in food animal. In the USA 80% of annual antimicrobial consumption is in food animals. The use of antimicrobials in animal feed in India is projected to increase by 82% between 2010 and 2030 and their use in chicken is expected to triple between 2010 and 2030 (Van Boeckel *et al.*, 2015). Tetracyclines accounted for the largest proportion of overall antimicrobial use globally (37.1% of total), followed by polypeptides (15.7%), penicillins (9.8%), macrolides (8.9%) and aminoglycosides (7.86%) (OIE, 2019).

Mechanism of development and spread of antimicrobial resistance

Several mechanisms are known for effecting antimicrobial resistance including through efflux pumps, selective porin channels or inactivating enzymes, and to take rapid advantage of beneficial mutations in binding sites and cellular structures. Not all resistance mechanisms are found in all species of bacteria; particular resistance determinants are found in particular bacteria. For example, resistance to macrolides is determined by different mechanisms in campylobacter compared with those in staphylococci. When antibiotics are administered for more than 10 days, bacteria may become resistant to that antibiotic as well as other antibiotics. As an example, 2 days after the administration of tetracycline to chickens, tetracycline resistant *E. coli* can be isolated from fecal samples, and within 2 weeks, *E. coli* becomes resistant

not only to tetracycline but also to multiple other drugs. Once the administration of antibiotics ceases, the loss of resistance is a slow process. Even after the antibiotic is removed from the environment, some bacteria populations can retain their antibiotic resistance for extended periods of time (Luo *et al.*, 2005). Gene mutation that confers resistance to some of the antibiotics e.g. fluoroquinolone confers a fitness advantage to the pathogen in the absence of antibiotic pressure and hence the resistance gene is retained for much longer period (Luo *et al.*, 2005). Resistance genes can spread through bacterial populations by different mechanisms like conjugal transfer of plasmids and other mobile genetic elements or by direct uptake of naked DNA (natural transformation) or by the action of bacteriophages (transduction).

Over time bacteria have been progressively resistant to multiple classes of antibiotics and they are called multi drug resistant (MDR) bacteria. Sometimes genes encoding resistance to one antibiotic in a class have the capacity to encode resistance to all antibiotics in that class and the phenomenon is called cross resistance. Another issue is co-selection for resistance (Cantón and Ruiz-Garbajosa, 2011). This occurs when multiple antibiotic resistance genes are located on a single plasmid or other mobile genetic element, so one of the co-located genes selects for resistance to all antibiotics on that plasmid or mobile genetic element. Generally it is accepted that the use of antimicrobials in human is the major driver of resistance development but, using antibiotics in animals also plays a role, though the data available to date are insufficient to quantify the contribution (Wall *et al.*, 2016). Most bacteria and their genes can move relatively easily

within and between humans, animals and the environment. Forslund *et al.* (2014) demonstrated that antibiotic use both in humans and in animals determines the resistance profile of bacteria in the human gut. The vast majority of antimicrobial classes are used both in humans and animals (including aquaculture; both farmed fish and shellfish). Only few antimicrobial classes are reserved exclusively for humans (e.g., carbapenems). There are also few classes limited to veterinary use (e.g., flavophospholipols, ionophores); mainly because of toxicity to humans. Insects (e.g., bees) and some plants are frequently treated with antimicrobials. Tetracyclines, streptomycin and some other antimicrobials are used for treatment and prophylaxis of bacterial infections of fruit, such as apples and pears.

In intensive animal farming and aquaculture, antimicrobials are often administered in sub therapeutic dose for prolonged period (two weeks and often almost the entire life of an animal, for example in chicken for 36 days or more) as growth promoter (FAO, 2018). These conditions favour selection and spread of resistant bacteria within animals and to humans through food or other environmental pathways.

Meat and egg from chickens reared under AGP may bear multidrug resistant zoonotic bacteria like *Campylobacter*, *E.coli* and *Salmonella*, and affect human health besides bearing AGP residues (Mehdi *et al.*, 2018). *Campylobacter* is a major cause of food-borne diarrheal diseases in humans. According to CSCRA (2016) report, chicken contamination rates for *E. coli*, *Campylobacter* and *Salmonella* spp. are respectively 96%, 25% and 34% in Canada. Antibigram test revealed

multi-pharmacological resistance in *Enterobacteriaceae* isolates from eggs and broiler meat (Diarra *et al.*, 2010; Singh *et al.*, 2010; Yulistiani *et al.*, 2017). In one study on *Salmonella enterica* isolates collected from poultry farms in British Columbia (Canada), Diarra *et al.* (2014) showed that more than 43% of the isolates were simultaneously resistant to ampicillin, amoxicillin-clavulanic acid, ceftiofur, cefoxitim and ceftriaxone. Another Canadian study (Diarra and Malouin, 2014) highlights the existence of different stereotypes of *Salmonella*, isolated from broiler farms, resistant and multi-resistant to antibiotics.

Large amounts of antibiotics administered to animals are excreted into the environment via urine and faeces. After metabolic changes in animals, 30% to 90% of the dose consumed is found in the urine and feces as parent compounds and/or metabolite compounds (Carvalho and Santos, 2016). This makes sewage disposal systems one of the most important routes by which antibiotics can enter in the environment and contaminate even coastal waters (Chen *et al.*, 2015). The aquatic environment is considered as an important point for acquisition and spread of antibiotic resistance genes by bacteria (Devarajan *et al.*, 2017). The soil can also be contaminated by antibiotics in litter. Animal bedding contains residues of antimicrobial compounds. Residues of bacitracin, salinomycin, penicillin and virginiamycin were detected in chicken litter at concentrations ranging from 0.07 to 66 mg/L (Furtula *et al.*, 2010). When this bedding material is used as nitrogen amendment, the resistant bacteria can live in the soil for several months (Merchant *et al.*, 2012).

Antibiotic used in aquaculture, apiculture and in plants or fruit crops, effluents from pharmaceutical industries and hospital waste are also being implicated for development of antimicrobial resistance.

Use of antibiotics in different ecosystem has led to environmental contamination (da Costa *et al.*, 2013). Human sewage, livestock production, and aquaculture have all contributed to widespread distribution of antibiotic-resistant bacteria and genes in the environment (Abhirosh *et al.*, 2011) facilitating their transmission to humans and animals.

Global use of antibiotic growth promoters in livestock and poultry and their linkage with antimicrobial resistance

Growth enhancement with antibiotics (sulphonamides) was first observed by Moore *et al.* (1946). The discovery by Jukes and his colleagues at Lederle Laboratories (Jukes, 1977) that aureomycin stimulated significant growth in chickens, cattle and pigs was the foundation of antibiotic growth promotion in animals.

About 40 percent of globally produced antibiotics are used as growth promoters (Hughes and Heritage, 2004). Antibiotic growth promoters (AGP) are commonly used in intensive cattle, pig and poultry farming to increase growth, improve feed efficiency and reduce disease incidence.

The use of antibiotics as animal growth promoters differs between countries. Sweden and UK now makes no use of antibiotics for growth promotion purposes (Begemann *et al.*, 2018). The U.S. has removed all growth promotion clearances for medically important antibiotics from 2017, but allow use of ionophores and non-medically important

prophylactic antibiotics (primarily bacitracin, avilamycin and bambermycins) (Smith, 2019). Virginiamycin was being used in different countries for prevention of lactic acidosis in cattle and as AGP in poultry, but use of this compound has led to the selection of bacteria that are resistant to its effects. It is related to pristinamycin and quinupristin, which are used in human medicine and so there are fears that its continued use may compromise human therapy. Its use as a growth promoter was banned in the EU. In 1995 vancomycin-resistant *Enterococcus faecium* was found in pigs and poultry that had been fed avoparcin-medicated feed (Bager *et al.*, 1997). In 1996 the EU suspended the registration of avoparcin. Apart from avoparcin there were also concerns about tylosin in co-selecting for macrolide (eg erythromycin used in human) resistance and co-selecting for avoparcin resistance (Butaye *et al.*, 1999). In 1999 EU suspended the use of tylosin, spiramycin, bacitracin and virginiamycin. By 2006 no AGPs remained registered in EU. Studies indicated that removing AGPs resulted in a substantial decline in antibiotic use (Millet and Maertens, 2011), had not increased the use of therapeutic antibiotics except in weaner pigs in Denmark (Grave *et al.*, 2006) and had led to a decline in antibiotic resistance in enterococci isolated from animals and food (Wegener, 2003). In Australia the use of gentamicin is prohibited, fluoroquinolone use in food-producing animals has never been approved, use of 3rd generation cephalosporins is restricted and cefquinome has not been registered (APVMA, 2017). In India, Food Safety and Standards Authority of India issues advisory regarding limits of antibiotic residues in animal products for most of the commonly used antibiotics and

Ministry of Health and Family Welfare and Ministry of Fisheries, Animal Husbandry and Dairying has banned use of colistin and its formulations for food producing animals, poultry, aqua farming and animal feed supplements in 2019 (Anonymous, 2019).

Antibiotics used for therapeutic purpose in animals linked to antibiotic resistance

Until quite recently only AGPs were being blamed for increased AMR instances. However, recently many antibiotics used in animals for therapeutic purpose are also being implicated for AMR. Enrofloxacin (a fluoroquinolone) was introduced in Europe and the USA in the late 1980s and resistance was first reported in veterinary isolates of *Salmonella* in the mid-1990s (Griggs *et al.*, 1994). Subsequent studies have confirmed the use of fluoroquinolones in food-producing animals to be a significant factor in the emergence of resistance in human infections (Moore *et al.*, 2006; Nelson *et al.*, 2007). Plasmid-borne fluoroquinolone resistance has now been found in animal isolates (de Jong *et al.*, 2014; Yanat *et al.*, 2017).

Ceftiofur, a 3rd generation cephalosporin, was introduced to the USA in 1998 and prior to that in Europe and strong links between the emergence of extended spectrum beta lactamases (ESBLs) mediated resistance to ceftriaxone in human and the use of ceftiofur (and cefquinome, which has been approved for use for some time in Europe but is not currently approved in the USA or Australia) in livestock has been recognized (Tyrrell *et al.*, 2016). The enzymes that attack the 3rd and 4th generation cephalosporins such as ceftiofur and cefquinome respectively are

now often referred to as Extended Spectrum Cephalosporinases (ESCs). There has been increase in the number of reports of ESC-resistant organisms with the introduction of cefquinome, a 4th generation cephalosporin, onto the EU market for use in dairy cattle and horses. ESC activity has been reported in *E.coli* and *Salmonella Enterobacteriaceae* including *Citrobacter* spp., *Klebsiella pneumoniae*, *Serratia* spp. and *Enterobacter cloacae* as well as non-*Enterobacteriaceae* such as *Acinetobacter* (APVMA, 2017). Except *E. Coli* and *Salmonella* these organisms have largely been ignored in veterinary testing because most of them are not considered as animal pathogens and role of animals in the dissemination of such non-*E.coli/Salmonella* MDR organisms is slowly being revealed (Müller *et al.*, 2014). However, antibiotic-resistant bacteria/genes could also be transferred from humans to animals (Schultz *et al.*, 2015).

Resistant animal pathogens with multi-resistance conjugative elements and multi-resistance genes which encodes resistance to phenicols, lincosamides, oxazolidinones (such as linezolid), pleuromutilins and streptogramin A, were reported in staphylococci of animal origin and other Gram-positive bacteria and in enterococci and animal isolates of Gram-negative bacteria such as *Proteus* and *E. coli* (Shen *et al.*, 2013; Tamang *et al.*, 2017).

Currently carbapenemase-producing and colistin-resistant MDR Gram-negative bacteria have been on focus as carbapenems and colistin are antibiotics of last resort for infections caused by MDR Gram-negative bacteria (Al-Tawfiq *et al.*, 2017; Madec *et al.*, 2017).

Carbapenems are not registered for use in animals but resistance has been detected in livestock, seafood, horses and companion animals (Madec *et al.*, 2017; Guerra *et al.*, 2014). The most likely cause for this is co-selection, though at first it was thought it might be the result of transfer from humans to animals or off-label or illegal use in animals (Falgenhauer *et al.*, 2017). Most of the ESC-producing organisms are multi-drug resistant and there are also reports of livestock and pet animals with ESC-resistant determinants on mobile plasmids (Seiffert *et al.*, 2013).

Methicillin-resistant *Staphylococcus aureus* (MRSA) first emerged in the 1960s and became the most common nosocomial pathogen in hospital patients (Moellering, 2012). Subsequently, there were increasing reports of MRSA in different animals (Sun *et al.*, 2015). The emergence of MRSA in humans led to introduction of vancomycin to treat resistant staphylococcal infections and this in turn led to the emergence of resistance to vancomycin.

Mechanism of action of antibiotic growth promoters

Healthy guts are essential to optimize digestibility in chicken. Intestinal epithelial cells are derived from cells situated in the intestinal crypt base. Cells proliferate by mitosis in the crypts, differentiating as they migrate upward to each villus and reach the villus tips, where they are shed into the intestinal lumen within 48 hours after birth (Potten, 1998). Generally fast growing broiler chicken are fed highly nutrient rich diets resulting in an excess of nutrients in the hindgut which causes the proliferation

of harmful microbes (bacteria, fungi, protozoa, viruses ,etc) in the hindgut with the consequence disruption of gut microbiome-host equilibrium (Weiss and Hennet, 2017) causing the metabolic, pathogenic or sterile inflammation of gut (Kogut *et al.*, 2018). Excess nutrients, especially protein and fat, are not well digested or absorbed which allows microbial proliferation in the ceca. With physiological retroperistalsis, these bacteria go back to the ileum and jejunum causing dysbacteriosis and even disease by the production of endo and exotoxins. Disturbances in the bacterial microbiota, due to this proliferation, result in dysregulation of adaptative immune cells and changes in microbial metabolism that can lead pathobiont microorganisms to become pathogens (Round and Mazmanian, 2009). AGPs have been the most consistent cost effective tool currently used to control dysbacteriosis and reducing microbial fermentation associated wastage of nutrients. Meta-genome sequencing approaches have demonstrated that different AGPs have different signature effects on gut microbiome. For example, populations of *Lactobacillus* spp. in the ileum of chickens receiving feed containing tylosin, a bacteriostatic, are significantly lower than those in chickens receiving no tylosin (Lin *et al.*, 2013). The decrease in the *Lactobacillus* population in antibiotic-treated animals probably reduces the intestinal activity of the bile hydrolase salts, which would increase the relative abundance of conjugated bile salts, thus promotes lipid metabolism and energy harvesting and increases animal weight gain (Lin *et al.*, 2013)

Besides suppressing sensitive populations of pathogenic or select non-pathogenic bacteria

in the intestines thus reducing microbial fermentation associated wastage of nutrients in intestine and by reducing pathogen load and toxins, antibiotics are also believed to play important role by lowering need for immune response including cytokine release which lower the release of catabolic hormones and in turn muscle wastage, reducing intestinal bacterial (*Lactobacillus* and *Bifidobacteria*, etc.) bile salt hydrolase activity thus increasing fat digestion, lowering gut epithelial, turnover, inflammation and thickness leading to better absorption of nutrients and incidence of bacterial diseases especially under poor hygienic condition and under heat or humidity stress. Whatever the mechanism of action is, the production benefits of the use of antibiotic growth promoters ranges between 1 and 10 percent. Prescott and Baggot (1993), however, showed that the effects of growth promoters were much more noticeable in sick animals and those housed in cramped, unhygienic conditions. Surveillance and animal production data however now suggests that benefits in animals reared in good conditions are probably quite small and may be non-existent.

Antimicrobial resistance status in India and national action plan

In 2017, the Centre for Science and the Environment, a non-profit organization based in New Delhi released a study entitled – ‘Antibiotic Resistance in Poultry Environment’ (Bhushan *et al.*, 2017). The study found that 100% of the *E. coli*, 92% of *K. pneumoniae* and 78% of *S. lentus* isolated from the poultry environment were multi-drug resistant. About 40% of *E. coli* and 30% of *K. pneumoniae* isolates were resistant to at

least 10 out of 13 antibiotics against which these bacteria were tested for resistance. Both *E. coli* and *K. pneumoniae* had very high resistance to antibiotics such as penicillins, fluoroquinolones, third and fourth generation cephalosporins and carbapenems, which is a last resort antibiotic used in hospital.

As part of the 'National Programme for Containment of AMR' (2012–2017), it was decided to establish a laboratory based surveillance system by strengthening laboratories for AMR in the country and to generate quality data on antimicrobial resistance for pathogens of public health importance. The ICMR's Antimicrobial Resistance Surveillance and Research Network (AMRSN) is currently carrying out surveillance with a network of 16 laboratories across the country (Walia *et al.*, 2019). A total of 30 labs in state medical colleges are planned to be strengthened in a phased manner to carry out surveillance. However, in the case of AMR in animals and food or antibiotics there is very limited surveillance. There are isolated studies, which have indicated high levels of AMR across animal commodities and systems, but they are yet to be unified under a nationally scaled programme. The Indian National Action Plan on AMR emphasizes a One Health approach.

In 2017, Indian health authorities released their National Action Plan on Antimicrobial Resistance (NAP-AMR) 2017–2021, that outlines the various challenges that need to be tackled to manage the phenomenon. There are six key areas that the NAP-AMR has identified as being strategic priorities for Indian health authorities to take action on. These include: Improved awareness of AMR through effective communication;

Strengthening knowledge and evidence through surveillance; Reducing the incidence of infection through effective infection prevention and control; Optimizing the use of antimicrobial agents in health, animals and food; Promoting investments for AMR activities, research and innovations; and Strengthening India's commitment and collaborations on AMR at international, national and sub-national levels.

Strategies for reducing use of antibiotic growth promoters

In developing countries like India antimicrobial growth promoters are often used to compensate for poor hygiene, housing and as replacement for proper animal health management. However, improvement in performance with AGP are in the range of 5-10% and such improvements are not very consistent and are largely limited to the period of high environmental stress. Keeping in view emerging threats of AMR for human and animal health, it has been advocated that use of antimicrobials as AGP should be banned. However, data on economic impact of such a blanket ban on Indian farmers has not been worked out. In the following section some of the strategies which are likely to contribute significantly in lowering use of AGPs in food animal rearing will be reviewed.

Use of different alternatives to antibiotic growth promoters

Research has been carried out to identify alternatives with similar beneficial effects of antibiotic growth promoters. Among these, the most popular are probiotics, prebiotics, enzymes, organic acids, immunostimulants, bacteriocins, bacteriophages, plant bioactive feed additives, phytoncides, nanoparticles

and essential oils. Probiotics introduce desirable microorganisms into the gut. However, microbes used as probiotics are not exempted from acquiring antibiotic resistance genes. Given their shared microbial environment in the gastrointestinal tract, a risk of pathogenic microbes acquiring antibiotic resistance genes from probiotic microbes exists, and vice versa. Hence, probiotics need to be screened for antibiotic resistance (Imperial and Ibana, 2016). Prebiotics promote the growth of desirable bacteria in the gut whereas organic acids cause the inhibition of harmful bacterial growth (Dittoe *et al.*, 2018). Enzymes such as phytases, carbohydrases and proteases have been widely used due to their cost saving impact on diet formulation as productivity enhancers and also for their positive impact on gut microbiome. Exogenous enzymes reduce microbial proliferation by reducing the undigestible components of feed, phytate, the viscosity of digesta or the irritation to the gut mucosa that results in inflammation. Enzymes also generate metabolites that promote microbial diversity which helps to maintain gut ecosystems that are more stable and more likely to inhibit pathogen proliferation (Kiarie *et al.*, 2013). The effects of plant bioactive compounds vary depending on their composition. They can be bacteriostatic or immune-stimulating. One category of plant bioactives are the essential oils (EO) or their active ingredients such as carvacrol, thymol, and cinnamaldehyde, etc., that have selective antimicrobial properties. The use of some specific EO blends has been shown to have efficacy towards reducing the colonization and proliferation of *Clostridium perfringens* and controlling coccidia infection and, consequently, may help to reduce necrotic enteritis (Oviedo-Rondón *et al.*, 2010). Some evidence suggests that the combined

administration of essential oils plus organic acids or prebiotics and probiotics might be the more effective than when used alone. Rearing chicken on these alternatives likely to be costlier than when raised on AGPs and performance may also be subpar.

Improving water and feed quality

Good water quality is also important for proper digestion and maintaining healthy gut. The pH of water should be maintained slightly acidic, between 5.5 and 7, because basic water reduces the activity of most digestive enzymes. For this reason, drinking water with high levels of carbonates or other salts that increase the alkalinity and hardness can cause problems. High concentrations of salts and solids in the water and basic pH generally favor the production of biofilm and endotoxins in water lines and drinkers due to the proliferation of algae and microbes. Unhealthy gut due to poor water quality may increase use of antibiotic and promote antibiotic resistance. When the pH value of the crop is above 6.5 the enzyme activity can be reduced 10–15% compared to the effectiveness observed at pH 4.5–5.5 (Kierończyk *et al.*, 2016). Organic acids in feed or water create acidic conditions in the crop of broilers, but not in layers. Dietary calcium levels higher than the requirement for each age group and productive phase can be deleterious to intestinal health since it may chelate nutrients, reduce enzyme activity, and promote proliferation of *Clostridium* bacteria (Paiva *et al.*, 2013).

Dietary fat for poultry should be free from rancidity. Diet should be free from mycotoxins. Quality control for soyabean meal is also important. Undercooked

soybeans have higher antitrypsin factor concentrations whereas overcooking decreases protein digestibility.

Improving bio security and hygiene and development of effective vaccines

There is an urgent need to train farmers on bio security measures. Also there is urgent need to develop more effective vaccines against viral and bacterial diseases, which will go in the long way to reduce use of AGPs. Notably, in one study on 64 farms in 9 European countries, the majority of pig operations experienced cost reductions for antibiotic treatments after *L. intracellularis* vaccination, even though not all farms were able to reduce their antibiotic use (Adam, 2009). The development of new safe and effective adjuvants or the combination of vaccines with immune modulators may be a promising strategy for overcoming limitations in vaccine efficacy, in particular for relatively short-lived species such as poultry. In case of poultry, it has been suggested that development of new or improved vaccines for *E Coli*, infectious

bursal disease virus, *Clostridium perfringens* type A, Eimeria and infectious bronchitis virus may reduce antibiotic use dramatically (Hoelzer *et al.*, 2018)

Bans on growth promoters within Europe led to little or no impact on livestock productivity and disease, including in poultry, yet changes in the management of farms such as improvements in hygiene and biosecurity, and associated costs are not available in public domain. These results are based on European production systems with high levels of bio security, and are difficult to extrapolate to current antimicrobial use and potential impacts of change in antimicrobial use on AMR and livestock feed systems in India. There is urgent need to undertake detail study and economic analysis of various alternatives to antibiotics before farmers can be persuaded to do away with AGPs. Further, development of reliable certification system and higher pricing of poultry products produced through alternate system and increasing consumer awareness can be a way forward.

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